using the primer pair 5'-CCTGGATCCGAAAG-TATAGCTTCTACCATTG-3' (SEQ ID NO:14) and 5'-TACATAAGCTTCTAGAT-GGCCAGAAAAGGTTCAGCA-3' (SEQ ID NO:15). The resulting 1562 bp fragment was digested with HindIII and BamHI, and ligated in the pMal-c vector. The C-terminal fragment (mSTAU-C) was amplified with the primer pair 5'-GGATGAATCCTATTAGTAGACTTGCAC-3' (SEQ ID NO:16) and 5'-TACATAAGC-TTCTAGATGGCCAGAAAAGGTTCAG-CA-3' (SEQ ID NO:22), digested with HindIII and cloned in the EagI* and HndIII sites of pMal-c. EagI* was created by filling in the cohesive ends of EagI-digested pMal-c vector using the Klenow fragment of DNA polymerase I. This fusion vector was then digested with SacI and EcoRI and the resulting fragment was subcloned in the pMal-stop vector to generate the mSTAU-RBD3 construct. The mSTAU-TBD construct was prepared by PCR using the primer pair 5'-GCTCTAGATTCAAAG-TTCCCCAGGC-GCAG-3' (SEQ ID NO:17) and 5'-TTTAAGCTTCTCAGA-GGGTCTAGT-GCGAG-3' (SEQ ID NO:18); the product was digested with XbaI and HindIII and cloned in the pMal-stop vector. mSTAU-RBD2 and mSTAU-RBD1 were constructed by first amplifying a fragment using the primer pair 5'-CAATGTATAAGCCCGTGGACCC-3' (SEQ ID NO:19) and 5'-AAAAAGCTTGTGCAAGTCTACTAATAGGATTCACC-3' (SEQ ID NO:20). The resulting product was digested with HindIII and cloned in the EagI* and HindIII sites of the pMal-stop vector. This vector was then used to purify the 398 bp PstI and HindIII fragment, which was subcloned in the pMAL-stop vector to generate the mSTAU-RBD2 construct. In the same way, the mSTAU-RBD1 vector was obtained by digestion with SmaI and StuI, followed by recircularization of the digestion product using T4 DNA ligase. The mSTAU-RBD4 was PCR amplified using the primer pair 5'-ATAGCCCGAGAGTTGTTG-3' (SEQ ID NO:21) and 5'-TACAT-AAGCTTCTAGATGGC-CAGAAAAGGTTCAGCA-3' (SEQ ID NO:22)."

Kindly replace pages 45, lines 13 to 24 by the following:

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"5'-TACATGTCGACTTCCTGCCA/GGGCTGCGGG-3' (SEQ ID NO:23) and 5'-TACAATCTAGATTATCAGCGGCCGCACCTCCCACACACAGACAT-3' (SEQ ID NO:24). The 3'-primer was synthesized with a NotI site just upstream from the stop codon allowing ligation of a NotI cassette containing either three copies of the HA-tag or the GFP sequence. The resulting fragment was cloned in Bluescript